

2023-01-12

Different Contribution of Olive Groves and Citrus Orchards to Soil Organic Carbon Sequestration: A Field Study in Four Sites in Crete, Greece

Sofoulaki, Eleni





MDPI

<http://hdl.handle.net/11728/12334>

Downloaded from HEPHAESTUS Repository, Neapolis University institutional repository

Article

Different Contribution of Olive Groves and Citrus Orchards to Soil Organic Carbon Sequestration: A Field Study in Four Sites in Crete, Greece

Eleni Sofoulaki ^{1,2}, Vasileios A. Tzanakakis ^{2,*}, Georgios Giannopoulos ³ , Iosif Kapellakis ⁴,
Emmanouil Kabourakis ² , Theodoris Chatzistathis ⁵  and Nikolaos Monokrousos ¹ 

¹ Department of Humanities Social Sciences and Economics, International Hellenic University, 57001 Thessaloniki, Greece

² Department of Agriculture, School of Agricultural Sciences, Hellenic Mediterranean University, 71410 Heraklion, Greece

³ School of Agriculture, Aristotle University of Thessaloniki, 54124 Thessaloniki, Greece

⁴ Department of Civil Engineering, School of Architecture, Engineering, Land and Environmental Sciences, Neapolis University, 2 Danais Av., 8042 Pafos, Cyprus

⁵ Institute of Soil and Water Resources, Hellenic Agricultural Organization (ELGO-Dimitra), 57001 Thessaloniki, Greece

* Correspondence: vtzanakakis@hmu.gr; Tel.: +30-6977441455

Abstract: In this work, we evaluated the effects of cultivation practices and sites (representing four locations in Crete, Greece) on soil organic carbon sequestration in established citrus orchards, olive groves, and uncultivated fields (used as a control). Soil pH, soil texture, soil organic matter (SOM), Permanganate Oxidizable Carbon (POXC), Total Kjeldahl Nitrogen (TKN), Carbon and Nitrogen ratio (C:N), as well as soil CO₂ respiration rates, and specific enzymes' activity (i.e., N-Acetyl Glutamate (NAG), Beta Glucosidase (BG), Dehydrogenase) were determined in the upper soil layer (0–20 cm). It was shown that citrus and olive orchards under the South Mediterranean conditions could substantially increase C storage in the soil. However, soils planted with orange trees showed lower capacity than olive trees, which was related to litter chemistry (i.e., leaf C:N ratio). Sites had no significant impact on SOM. In our study, SOM had a positive relationship with TKN (and less with POXC) and the C:N ratio of the tree crop species litter. Our findings have implications for designing soil conservation practices in Mediterranean conditions and developing initiatives describing achievable targets of SOM restoration depending on soil properties and cropping systems.

Keywords: organic C sequestration; olive groves; citrus orchards; SOM; POXC; Mediterranean agroecosystems; adaptation and mitigation of climate change



Citation: Sofoulaki, E.; Tzanakakis, V.A.; Giannopoulos, G.; Kapellakis, I.; Kabourakis, E.; Chatzistathis, T.; Monokrousos, N. Different Contribution of Olive Groves and Citrus Orchards to Soil Organic Carbon Sequestration: A Field Study in Four Sites in Crete, Greece. *Sustainability* **2023**, *15*, 1477. <https://doi.org/10.3390/su15021477>

Academic Editor: Jeroen Meersmans

Received: 2 November 2022

Revised: 7 January 2023

Accepted: 10 January 2023

Published: 12 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Protection and enhancement of stored carbon (C) in the soil is an important goal for sustainable natural resource management to preserve the quality and health of soils and to sustain ecosystem services [1]. This target is essential for arid or semi-arid areas, such as the Mediterranean basin, due to environmental and climate conditions (e.g., high temperature and evapotranspiration, low precipitation rates, and drought events) and intensive management practices and constraints on litter inputs to the soil (e.g., tillage, high inorganic fertilization, and residue burning) [2]. To date, several studies have provided evidence for a low potential of Mediterranean agroecosystems, especially in its southern part, to store C, even under soil conservation practices (e.g., non-tillage), questioning the capacity of commonly applied practices to restore soil health, mitigate climate change, and improve the resilience of agroecosystems to climate extremes. Therefore, more efforts should be made to adapt crops and agricultural practices and to respond to current economic, environmental and climate challenges [3,4].

Plant species are essential to agriculture and food production, determining crop production, cropping systems' environmental and climate footprints, and total C sequestration [5]. They balance the direct or indirect (via plant-regulated soil microorganisms activity) CO₂ emissions with the C stored in plant biomass and the soil (C sequestration) [6]. Tree plantations are featured by their capability to absorb CO₂ from the atmosphere and temporarily store it over long periods within tree counterparts, such as branches, roots, trunks, fruits, leaves, and in the soil, and via litter decomposition and rhizodeposition [6–9]. The soil C sequestration is heavily regulated by critical factors and their interaction, such as crop-tree and soil characteristics, the farming system (e.g., conventional/organic), environmental conditions, and the applied agricultural practices (e.g., crop rotation, fertilization management, irrigation, tillage system, and residue management) [10–14].

Olive (*Olea europaea* L.) is the most economically and ecologically important tree crop in the Mediterranean area [15]. Related works in olive tree plantations provide information concerning the net C concentration [16,17], C exchange, and water use productiveness in irrigated olive fields [18]. Furthermore, models are suggested for calculating the growth potential of an olive tree's aboveground parts to measure C seizure [19]. Examining olive groves in Southern Italy, Sofo et al. [17] determined the average volume of stored CO₂ estimated as a function of belowground and aboveground structure differences, olive tree density, and age, and the results demonstrated values of 9.54 and 2.74 Mg ha^{−1} year^{−1} in mature and young groves, respectively. Lopez et al. [20] claimed that olive trees could stably store carbon, especially in intensive plantations, so olive groves decrease greenhouse gas emissions of greenhouse gases, highlighting their role as CO₂ sinks. They also reported that a reduced percentage of soil organic matter in intensive plantations was due to the inadequate management of the soil. Differences in soil C sequestration have been shown between olive orchards and avocado trees planted in a semi-arid region of Mediterranean, in Crete, which was attributed to crop litter characteristics [7].

Orange tree orchards are widespread worldwide, with the countries around the Mediterranean basin constituting important producers [21]. Unfortunately, the Mediterranean climate exhibits scarce water resources that confine orange crop sustainability [21]. In regions exhibiting a semi-arid climate, such as Crete, limiting water availability is the aspect that mainly contributes to the degradation of crop production [22]. Therefore, it becomes significant to settle the timing and amount of water to be irrigated [23]. Despite the sufficient number of studies conducted with orange trees, most focused on irrigation's impacts on physiology and productivity [24,25].

The island of Crete, due to its geographical location, is considered vulnerable to climatic conditions; it will be challenged in the upcoming years by climate change and/or variability [2,26] and water scarcity phenomena, particularly in the eastern-south part of the island [27]. Olive tree crop species occupy the largest part of the island's agricultural land. Other tree cultivations, such as orange orchards or avocados, and vegetables are cultivated and co-exist with the olives. Allen et al. [28], examining fields in Crete, observed fields of both intensive and traditional types of olive cultivation. In more upland and marginal areas, they observed traditional terraced olive fields, some of which have been abandoned. Today we are noticing intensive and conventional types of olive cultivation in both categories' areas, revealing the progress in the agriculture sector that was recorded during the elapsed years (2006–2022). The same observations are made for orange cultivation. Both crops studied here are considered as important cropping systems for the economy of Crete [27].

Despite those mentioned above and the available knowledge on olive and orange orchards' crop productivity and economy, there is limited knowledge regarding their potential to preserve and store C in soil. Therefore, more research must be conducted to estimate the CO₂ removal capacity concerning plant biomass sequestered C for an examined tree system and the C concentration in the soil. Several studies have evaluated the production and net C budget in forests. Still, limited research has targeted crops, partly due to the raised uncertainties and difficulties in calculating farmlands' C volume [29–32].

Based on the above four different sites of Crete (characterized by different soil-climatic conditions), non-cultivated or planted Olive and Orange trees were investigated with the objectives: (a) to study the effect of different tree crop species (olives vs. citrus) and sites on soil organic C sequestration (C storage as soil organic matter (SOM)); (b) to identify the critical soil and plant-specific properties driving soil organic C sequestration; and (c) to provide additional information for the development of necessary adaptation measures in agriculture due to climate change and/or variability in semi-arid and climate vulnerable areas of the planet (e.g., the Mediterranean basin). The underlying hypotheses of the present study are the followings:

- Farming land use either with olive or citrus trees will increase the soil organic C potential due to litter fall and rhizodeposition (will be indicated as a difference between tree crop species cultivations and non-cultivated fields in four different sites).
- Different tree crop species will have different soil organic C sequestration potentials due to their distinct physiological and morphological characteristics (e.g., litter quantity and quality and root distribution and rhizodeposition) and expected crop-specific soil microbiota communities and activities involved in C and nutrient cycling in the soil.
- Sites will affect the storage of soil organic C due to site-specific adjustment of crops due to existing soil-climatic conditions and site-specific soil microorganisms and their responses to environmental conditions.
- Soil organic C concentration could be correlated with specific soil properties and plant species characteristics (i.e., litter C:N ratio). Both the latter factors could affect organic C retention and availability and shape soil microbiota, consequently impacting the SOM storage in the soil.

2. Materials and Methods

2.1. Site Descriptions

The selected sampling sites are located a) in the north and central part of Crete: Sisses (SIS), 35°24'04" N, 24°52'15" E (Altitude: 40 m) and Heraklion [Hellenic Mediterranean University (HMU), 35°19'02" N (Altitude: 11 m), 25°06'26" E and b) in the south and eastern part of Crete: Messara/(MES), 35°04'39" N, 24°54'37" E (Altitude: 294 m) and Sitia (SIT), 35°12'12" N, 26°02'11" E (Altitude: 252 m) (Figure 1). The average maximum and minimum month temperature for the different site areas are from 7 to 22 °C and from 20 to 31 °C for the winter and summer periods, respectively. The MES and SIT sites are among the areas with the highest maximum temperatures and lowest precipitation rates on the island, exhibiting an increased risk regarding drought vulnerability, climate change impacts, and desertification potential [33]. Rainfall is varied from ~650 mm in Heraklion (HMU and SIS) to a lower level of ~500 (MES) and 450 (SIT) mm [34]. The soils in SIT and MES are threatened with chemical and physical deterioration due to the intensive agricultural practices, overgrazing, and erosion.

In each sampling site, three paired fields, an uncultivated one, a second one planted with olive trees, and a third planted with citrus trees of the same age, were surveyed to examine the variability of SOM in the soil of different cropping systems. The size of the fields was the average size in Crete, 0.5–1.0 ha. Olives and citrus trees were 25–30 years old, grown with mild-conventional management practices, applying either low quantity of synthetic chemical fertilizers (i.e., <80 NPK kg ha⁻¹ yearly) or low organic additions (HMU), both combined with minimum irrigation and tillage practices. The soil texture of the sites is depicted in Table 1.



Figure 1. The locations of four sites included in the study in Crete island, Greece (Source: Google Earth).

Table 1. Soil texture analysis of the field sites of the study.

		Silt (%)	Clay (%)	Sand (%)	Soil Texture	CaCO ₃ (%)
HMU	Non-cultivated	71	3	26	Silt Loam	14
	Olive trees	75	1	25	Silt Loam	14
	Citrus trees	74	3	23	Silt Loam	13
MES	Non-cultivated	53	13	35	Silt Loam	17
	Olive trees	55	4	41	Silt Loam	18
	Citrus trees	47	14	39	Silt Loam	14
SIT	Non-cultivated	30	14	57	Sandy Loam	<1
	Olive trees	23	20	57	Sandy Loam	<1
	Citrus trees	38	13	48	Sandy Loam	<1
SIS	Non-cultivated	33	8	59	Sandy Loam	<1
	Olive trees	41	7	51	Sandy Loam	<1
	Citrus trees	36	9	55	Sandy Loam	<1

2.2. Field Samplings and Analyses

From each of the above experimental fields, we selected three representative trees, from where three respective composite soil core samples were collected. Each of the latter composite samples contained four cross samples (four samples per tree). Thus, three replicates were used. In total, thirty-six soil samples were collected from four different productive and diverse areas in Crete, SIT, HMU, MES, and SIS [4 sites \times 3 fields (olive, orange, uncultivated) \times 3 (Replicates-Stations) = 36 soil samples]. Samples were taken from the first 20 cm of the soil in November 2021. The selection of single sampling was considered adequate given the reasonably long time since the establishment of the crops that ensure adequate cumulative impact on soil organic C sequestration. Soil samples were transferred to the Soil laboratory at HMU. They were sieved through a 2 mm mesh, placed separately in transparent bags, and stored in the refrigerator at 4 °C for chemical and physical analyses.

Besides collecting soil samples, twenty-four leaves were collected randomly from each of the above trees in the olive and citrus orchards. The leaf samples were transferred to the soil laboratory, washed gently, and placed on filter paper. After drying, each sample was milled separately to create a soft powder. Next, each sample was put in paper bags and then placed in a dryer until the day of the analyses.

Particle size analysis for soil samples was carried out by the Bouyoucos soil physical-chemical analyses method [35]. The SOM was determined by the Walkley–Black method [36]. Total Kjeldahl Nitrogen (TKN) contents were measured according to the Kjeldahl method [37]. For each sample, 1 gr of soil and leaf tissue was digested with 20 mL sulfuric concentrated acid (H₂SO₄). After that, nitrogen content was determined by alkaline reduction with

sodium hydroxide (NaOH) and following titration with sulfuric acid (0.01 and 0.1 N for soil and leaf tissue, respectively). Active carbon was determined by Permanganate Oxidizable Carbon (POXC) method [38]. In 2.5 g. of each soil sample, added 2 mL KMnO_4 and 18 mL of deionized water. After 2 min shaking and 10 min incubation in a dark room, samples were diluted in 49.5 mL deionized water, and the absorbance was determined by using Shimadzu Spectrophotometer. Free CaCO_3 was determined by using Bernard Calcimeter. The activity of N-acetyl-glucosaminidase (NAG) was measured colorimetrically by 96-well microplates modified method [39]. Specifically, fresh soil, equivalent to 0.5 g dry weight, was added with substrate solution p-nitrophenyl- β -N acetylglucosaminide in acetate buffer (pH 5) and incubated for three h at 21 °C. After the p-nitrophenol reaction, the final product was measured colorimetrically at 405 nm, and the NAG activity was determined as $\mu\text{mol pNP g}^{-1} \text{ dry soil h}^{-1}$. Dehydrogenase activity and β -glucosidase (BG) were determined by soil analysis methods [40,41]. Dehydrogenase activity was determined via soil incubation supplemented with an aqueous 2,3,5-triphenyltetrazolium chloride solution and CaCO_3 . The incubation lasted 24 h at 37 °C, and the product (triphenyl formazan (TPF)) was measured colorimetrically (formazan release method). In addition, the activity of BG was determined according to Kapagianni et al. [41], expressed as $\mu\text{mol pNP g}^{-1} \text{ soil h}^{-1}$.

To determine respiration rates, soil subsamples from the above treatments equivalent to 50 g were placed into pint-size glass canning jars (10 cm \times 1.5 cm), sealed by a gas-tight screw lid. The CO_2 emitted from the pots was trapped in a plastic vial containing 10 mL of 1 M NaOH, placed inside the jar. CO_2 was determined by adding 1 mL of 1 M BaCl_2 into CO_2 traps and titrating to neutral pH with 1 M HCl. Sampling was carried out in triplicate at each time interval (2 to 7-day sampling was carried out) for all treatments, lasting 39 days. Soil incubations were performed under controlled conditions at 25 °C in the dark.

Total organic carbon was determined by weighted 0.01 g of leaf tissue and using a TOC analyzer, solid sample module. First, total Kjeldahl Nitrogen (TKN) contents were measured according to the Kjeldahl method assay. Next, 1 g of plant tissue was weighed, placed in a capsule, and transferred to the desiccator for measuring total organic carbon with a SHIMADZU Total Organic Carbon Analyzer using the solid sample module (SSM-5000A).

2.3. Statistical Analysis

To determine the effect of sampling site, management type (cultivation) (independent variables), and their interaction on the soil and leaf physical, chemical, and biological variables (dependent variables), a two-way analysis of variance (ANOVA) was used followed by a Fischer's LSD at $p < 0.05$. Furthermore, a Pearson correlation analysis was performed to examine the relationships between soil and leaf variables. To further explore whether sampling site or cultivation type is more important for the variability shown by the data, a principal component analysis (PCA) was conducted based on soil physicochemical and biological variables. Before analyses, the data were transformed appropriately when necessary to meet the assumptions of the ANOVA. Statistical analyses were performed by STATISTICA 9 software (Ver. 9).

3. Results

3.1. Soil Organic Matter (SOM), Permanganate Oxidizable Carbon (POXC), Total Kjeldahl Nitrogen (TKN), C:N Ratio, and pH

Soil organic matter (SOM) showed variation across different sites ($p < 0.05$) and cultivation ($p < 0.001$) (Figure 2). Olive orchards tended to have higher SOM compared with the orange orchards in all sites (HMU, MES, SIT, SIS) (Figure 2). Lower SOM values were found in uncultivated fields suggesting plant species' critical contribution to SOM enhancement. Different impacts on POXC were detected among sites ($p < 0.001$) and tree

crop species ($p < 0.001$) without a clear trend (Figure 3). Similar to SOM, uncultivated fields showed lower POXC values than tree cultivations (Figure 3).

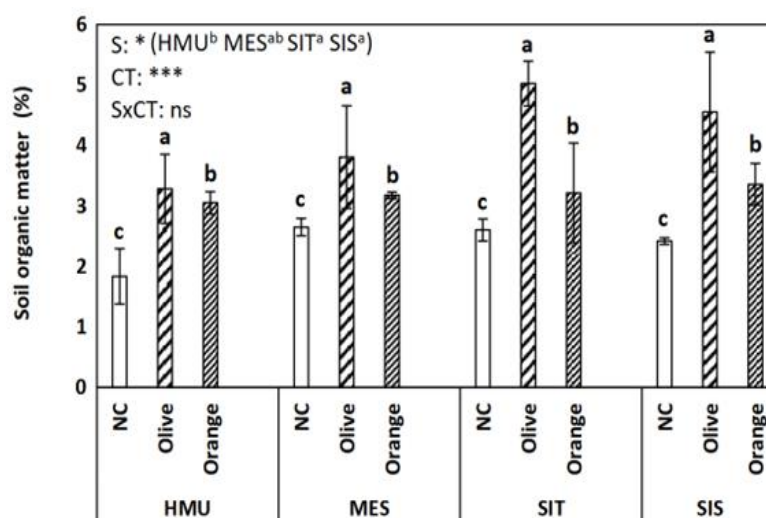


Figure 2. Mean values (\pm SE) of Soil organic matter (SOM) concentration in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (*: $p < 0.05$; ***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b,c” points to the highest value.

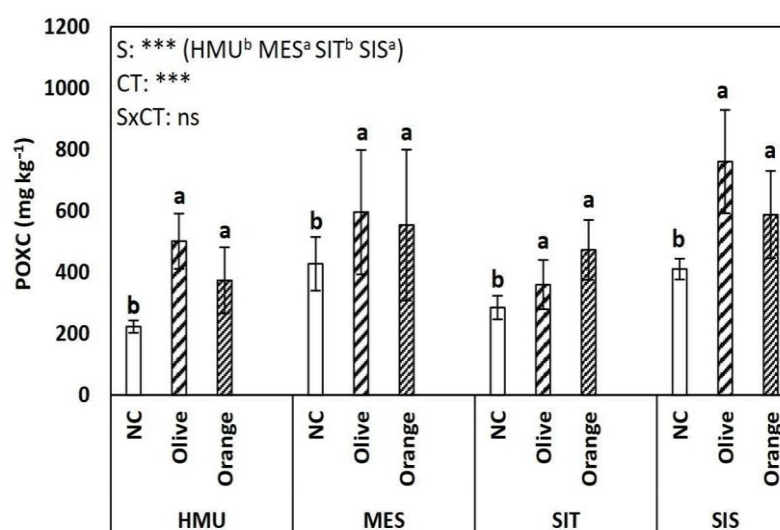


Figure 3. Mean values (\pm SE) of Permanganate oxidizable carbon (POXC) concentration in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b” points to the highest value.

Similar trend contrasts were also recorded for the parameters of the Total Nitrogen (TKN) (Figure 4) and C:N ratio (Figure 5), revealing an influence of site on olive and orange crops. Concerning the TKN, significant differences were observed only among crops ($p < 0.001$). Higher TKN values are recorded for fields planted with olive and orange compared with uncultivated areas (Figure 4).

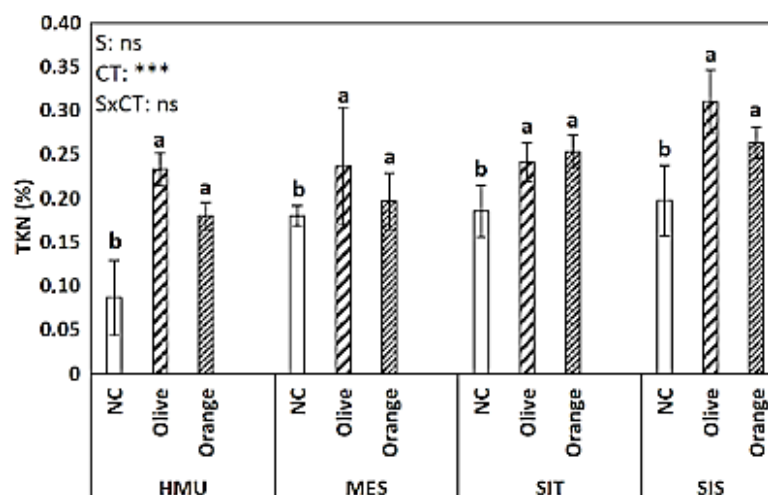


Figure 4. Mean values (\pm SE) of Total Kjeldahl Nitrogen (TKN) concentration in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (**: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b” points to the highest value.

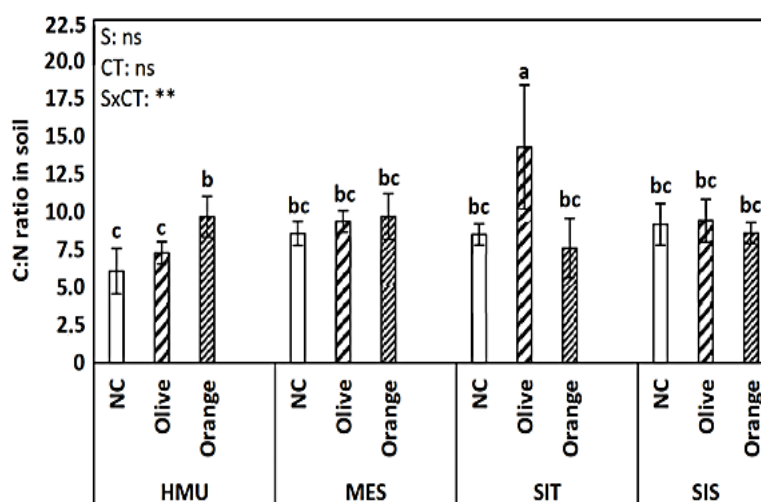


Figure 5. Mean values (\pm SE) of C:N ratio in the soil in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (**: $p < 0.01$; ns: non-significant, for all cases $n = 5$). The letters “a,b,c” points to the highest value.

The C:N ratio varied between sites and crops (Figure 5) and was influenced by site and tree crop species. The C:N ratio values in the cultivated and uncultivated fields in MES and SIS sites did not differentiate; on the contrary, in HMU, the highest C:N values were higher in the citrus orchard, whereas in the SIT site, the olive orchard was the one exhibiting the highest values.

The mean pH value in the uncultivated fields ranged from 6.5 to 7.8 (Figure 6). Furthermore, the pH factor was unaffected by the site or tree crop species. The olive groves always exhibited lower pH than the uncultivated fields (5–12% lower pH values). On the other hand, the citrus orchard pH values were always slightly higher than in olive groves, around 6–9%.

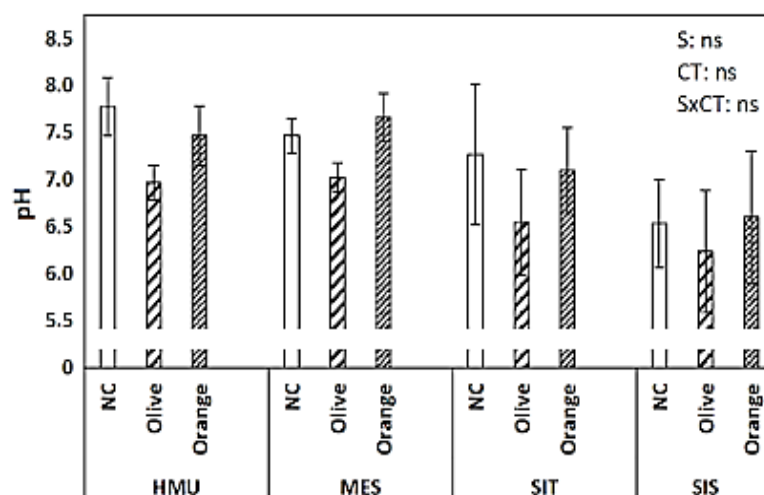


Figure 6. Mean values (\pm SE) of pH in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). (ns: non-significant, for all cases $n = 5$).

3.2. Respiration rates

The mean respiration rate ranged between 9.7 and 12.7 $\text{mg CO}_2 \text{ kg}^{-1} \text{ soil d}^{-1}$ in the uncultivated fields, 9.0 and 11.9 $\text{mg CO}_2 \text{ kg}^{-1} \text{ soil d}^{-1}$ in the olive groves, and 10.0 and 12.0 $\text{mg CO}_2 \text{ kg}^{-1} \text{ soil d}^{-1}$ in the citrus orchards (Figure 7). Therefore, there was no clear trend for this individual parameter.

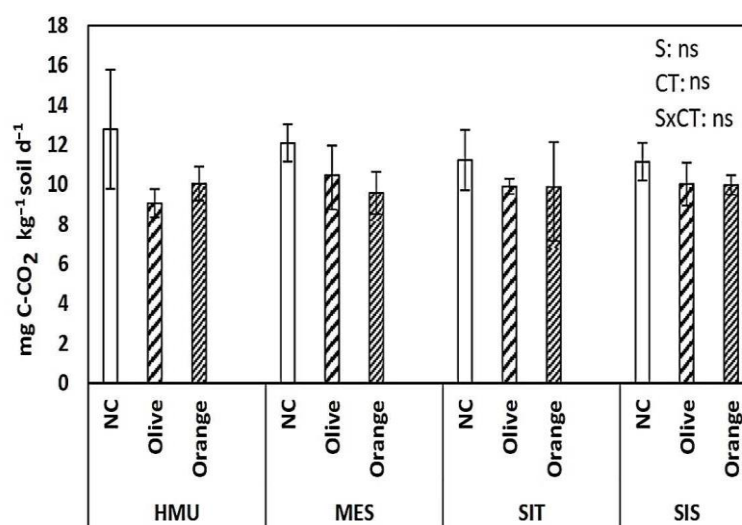


Figure 7. Mean values (\pm SE) of respiration rates (CO_2) in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). (ns: non-significant, for all cases $n = 5$).

3.3. Enzyme Activities

NAG activity did not exhibit a clear trend (Figure 8). In HMU, the uncultivated field had the highest values; in MES and SIT, the orange orchards showed the highest values, while in SIS, they were higher in olive cultivation. BG activity was significantly affected by the cropping system ($p < 0.001$) and site ($p < 0.001$) (Figure 9). The uncultivated fields had relatively low values. Similar values were recorded in olive and orange orchards (of around $1.2\text{--}1.3 \mu\text{molpNP g}^{-1} \text{ h}^{-1}$). Dehydrogenase activity was significantly affected by tree crop species ($p < 0.001$). The higher values were observed in the orange orchards,

followed by the olive orchards and the uncultivated fields. Sites did not significantly affect the dehydrogenase activity (Figure 10).

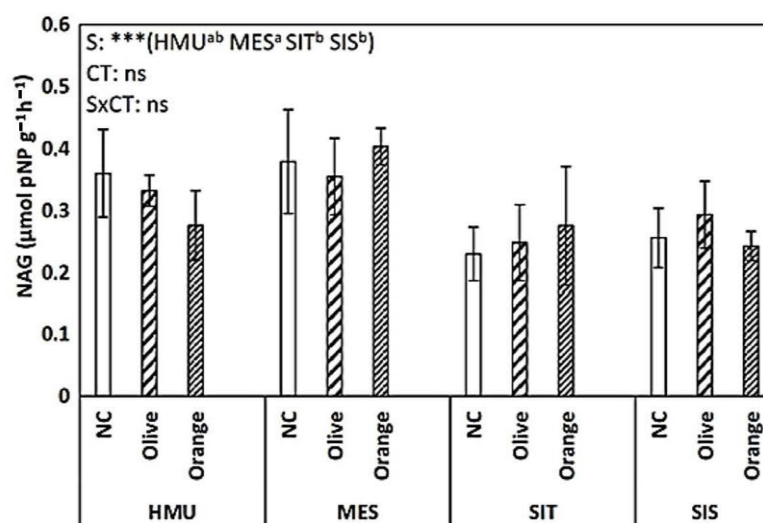


Figure 8. Mean values (\pm SE) of NAG activity in the soil in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b” points to the highest value.

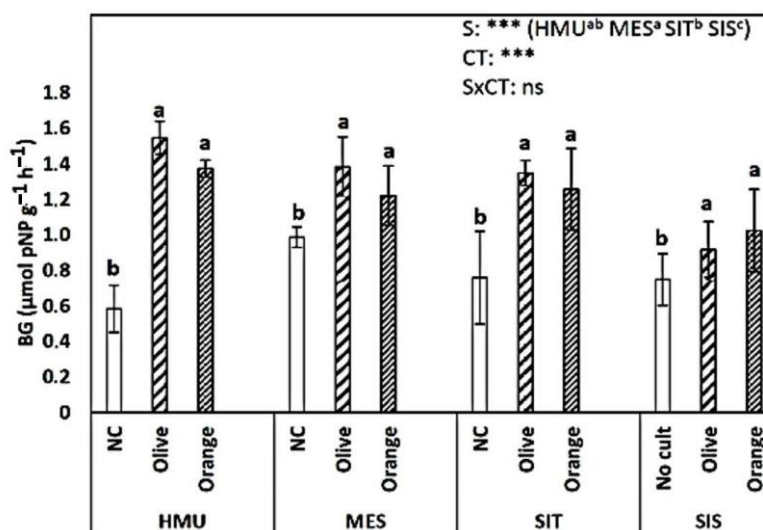


Figure 9. Mean values (\pm SE) of b-glucosidase activity (BG) in the soil in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b,c” points to the highest value.

3.4. Carbon, Nitrogen, and C:N Content in Leaves

Orange tree leaves tended to have higher leaf N content than olive trees, particularly in HMU and MES sites (Figure 11). However, the opposite trend was observed for these sites’ leaf C content, resulting in a significantly higher C:N ratio in olive than orange trees (Figure 11).

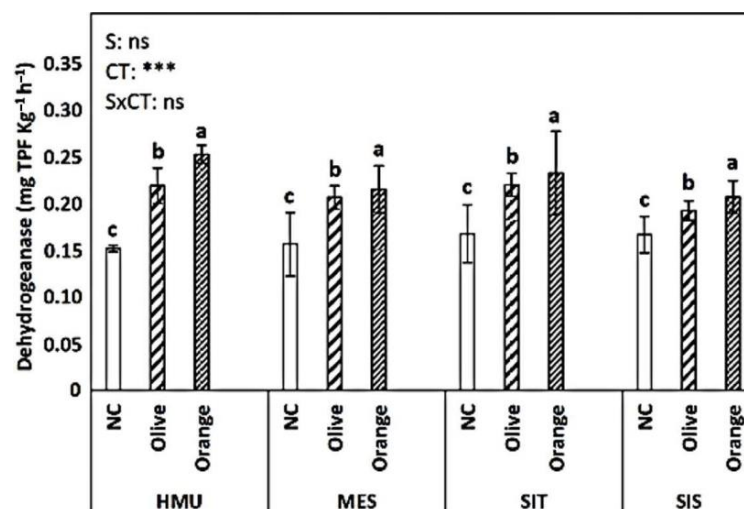


Figure 10. Mean values (\pm SE) of Dehydrogenase in the soil in different sites and crops (Non-cultivated (NC) vs. Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b,c” points to the highest value.

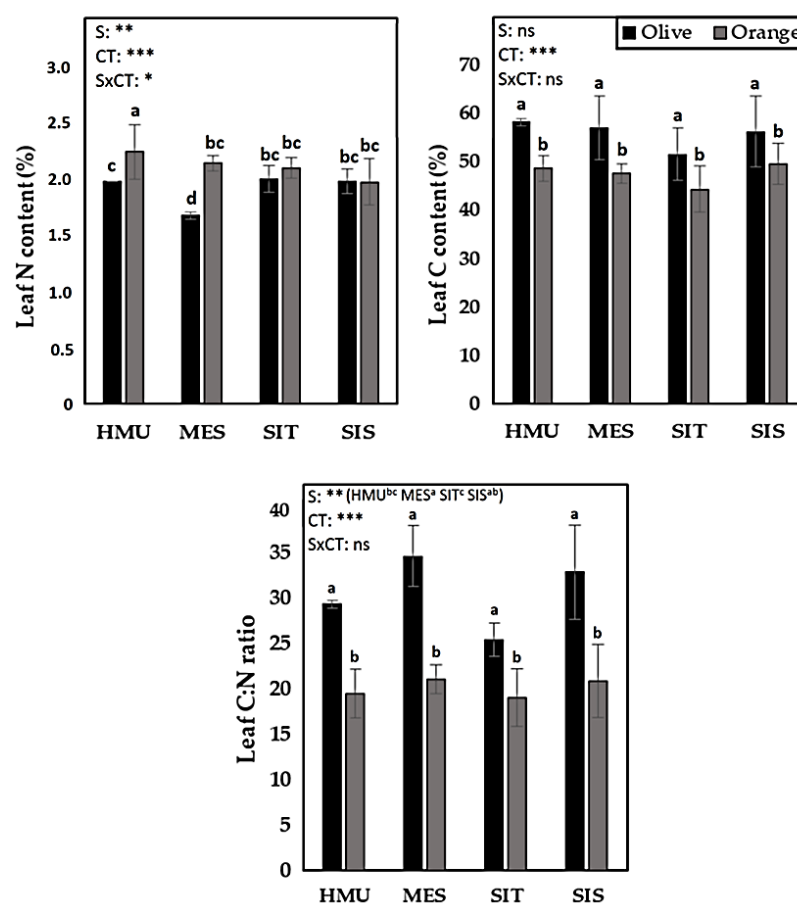


Figure 11. Mean values (\pm SE) of Leaf N, C, and C:N ratio contents in different sites and crops (Olive vs. Orange tree), and results of ANOVA regarding “Site” (S), “Crop Type” (CT), and their interactive effect (SxCT). Different letters indicate significant differences based on Fisher’s LSD post hoc test (*: $p < 0.5$; **: $p < 0.01$; ***: $p < 0.001$; ns: non-significant, for all cases $n = 5$). The letters “a,b,c” points to the highest value.

3.5. PCA and Correlation Analyses

The ordination of soil samples and variables on the PCA biplot is depicted in Figure 12. The first two PCA axes accounted for 60.04% of the data variability. The differentiation of samples due to the type of cultivation is shown along the first axis, which explained 39.69% of data variability. Samples from the uncultivated fields were ordinated within the left side of the biplot, no matter the sampling site, showing a positive correlation with CO₂; the right side was occupied by samples from the olive and orange cultivations (and SOM, TN, POXC, and DHG determined their distribution). The ordination of samples along the second axis reflected the site effect since the samples from HMU and MES occupied the lower part of the biplot and were characterized by increased values of silt, NAG, BG, and pH, while those of SIT and SIS were ordinated towards the upper side. The fact that the second axis explained only 20.35% of total data variability indicates that the cultivation effect is much stronger than that of the site on alterations of the soil's C-related properties.

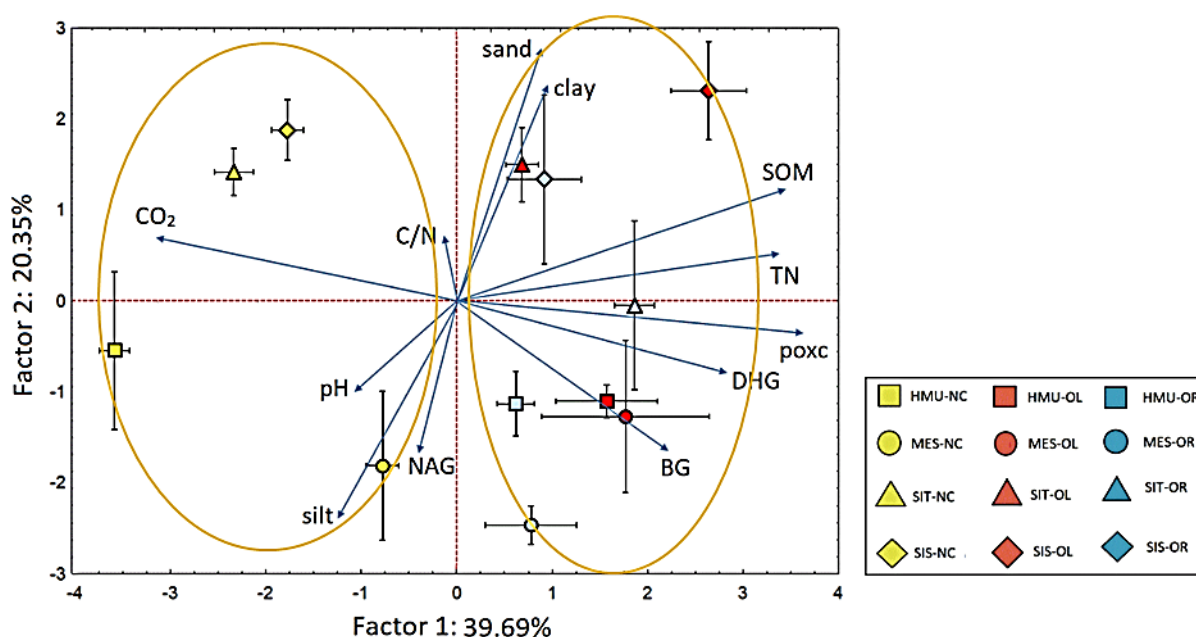


Figure 12. Ordination of soil samples based on soil physicochemical variables on a PCA biplot. Error bars indicate standard errors ($n = 3$). The first symbol indicates the sampling site, and the second is the crop type (NC: no cultivation; OL: olive; OR: orange).

Table 2 summarizes the main correlations between the various soil parameters examined. SOM showed a significant positive correlation with sand content and the TKN, soil, and leaf C:N ratio and a weak positive relationship with POXC; it negatively correlated with silt content BG. The latter enzyme activity showed a significant positive correlation ($p < 0.05$) with NAG and silt content. The CO₂ rate did not show any significant correlation.

Table 2. Correlation analysis for the soil and plant parameters examined in the present study.

	Silt	Clay	Sand	NAG	BG	DHG	POXC	TKN	pH	SOM	C:N	Leaf N	Leaf C	Leaf C:N
Silt														
Clay	−0.82													
Sand	−0.94	0.58												
NAG	0.43	−0.18	−0.50											
BG	0.43	−0.24	−0.47	0.41										
DHG	0.39	−0.17	−0.46	−0.09	0.31									
POXC	−0.15	0.01	0.21	0.16	−0.35	−0.37								
TKN	−0.07	−0.23	0.24	−0.20	−0.37	−0.28	0.49							

Table 2. Cont.

	Silt	Clay	Sand	NAG	BG	DHG	POXC	TKN	pH	SOM	C:N	Leaf N	Leaf C	Leaf C:N
pH	0.30	−0.11	−0.36	0.39	0.28	0.10	0.18	−0.11						
SOM	−0.44	0.34	0.43	−0.30	−0.44	−0.36	0.32	0.43	−0.22					
C:N	−0.41	0.55	0.26	−0.19	−0.03	−0.11	−0.14	−0.36	0.12	0.52				
Leaf N	0.04	0.22	−0.20	−0.14	−0.01	0.52	−0.10	−0.35	0.28	−0.16	0.18			
Leaf C	0.15	−0.31	−0.03	0.11	0.08	−0.47	0.12	0.23	−0.22	0.37	−0.02	−0.65		
Leaf C:N	0.10	−0.33	0.05	0.17	0.08	−0.53	0.12	0.29	−0.25	0.30	−0.10	−0.87	0.93	
CO ₂	0.11	−0.23	−0.02	0.00	−0.03	0.06	−0.20	−0.11	−0.07	0.05	−0.13	−0.09	0.17	0.19

Bold numbers represent significant correlation ($p < 0.05$).

4. Discussion

4.1. Tree Crop Species Contributed to Organic C Sequestration as Compared to Uncultivated Fields

The overall (above that found in uncultivated fields) contribution to SOC by tree crop species was estimated and the results varied between 2.4 and 15.7 t C ha^{−1}. We assume that organic C addition in the soil is mainly attributed to litter incorporation into the soil and subsequent SOM formation mechanisms and processes [42] involving rhizodeposition and soil microbiota assembly and activity [43–45]. Litter characteristics are important factors for C sequestration, including mainly the recalcitrant fraction of leaves and the C/N ratio [46–48]. The dominance of recalcitrant compounds in plant residues (i.e., lignin) may induce C stabilization in the soil via resistance to microbial decomposition and/or via interaction with the soil minerals, organic and inorganic compounds, environmental conditions, and soil microbiota [49–51].

Model-based simulations provide different estimates regarding the required amounts of C additions to restore or increase soil organic C in olive groves and citrus orchards. For example, there are CAST model-based simulations of SOM under different organic C rates, showing long-time C application to restore SOM, up to 12 t C ha^{−1} year^{−1} [52]. In a previous simulation study, organic inputs up to 25 t C ha^{−1} year^{−1} led to increases of SOC up to 14 t C ha^{−1} year^{−1} caused by changes in soil management from conventional tillage to mulching with residues from pruning debris and olive-fruit cleaning [53]. These values are higher than those of our study and are probably due to constraints from climatic conditions and the applied agronomic practices in the olive orchards of our research such as the low organic inputs and the pruning residues burning, a practice often used in olive orchards in Crete. Regarding citrus trees, a 7-year field study showed that SOC stocks increased by 38.6% under the combination of no-tillage and pruning residue (branches and leaves) incorporation or by 82.3% under reduced tillage, pruning residues incorporation plus drip-irrigation system compared with fields with intensive tillage and flood irrigation system, at 0–5 cm soil depth [42].

4.2. Different Contributions to SOM Enhancement between Different Tree Crop Species

The tree cropping system strongly affected SOM; olive groves achieved higher SOM values than the orange orchards. This finding may reflect the different litter loading, as well as the differentiations in litter quality (physical and chemical characteristics) between species. This hypothesis was confirmed by correlation statistics showing a strong positive relationship between SOM and leaf C, N, and the C:N ratio (Table 2) implying that the higher the C:N content in leaves the higher the potential for the soil C sequestration. Thus, the higher C:N ratio in olive than orange leaves may explain the higher SOM values in the former.

Earlier studies also highlighted the role of litter chemistry (e.g., cellulose, hemicellulose, and lignin content, as well as C and N content or the C:N ratio) on C mineralization and sequestration [46–48]. The C:N ratio increases have been related to increased microbial N immobilization and lower rates of C mineralization [54]. A recent study in Crete dealing with soil C sequestration in olive and avocado orchards reported different performances

between olive groves and avocado plantations, mainly attributed to different litter inputs and litter chemistry [7].

4.3. Limited Effect of Site Position on Soil Organic C Sequestration

According to our findings, sites had a limited weaker impact on SOM. We speculate that tree crop species, through their strong impact on SOM, have masked or mediated the influence of other biotic and abiotic parameters driven by field sites, such as site-specific soil biological and physicochemical properties, environmental conditions (e.g., precipitation and temperature), and their interactions [55–57]. Another explanation may include the synergetic influence of various biotic and abiotic factors on SOM impeding the extraction of clear results. It is well documented that the role of soil properties on microbial accessibility to organic substrates influences organic matter decomposition and interaction with other parameters [58]. Moreover, different environmental conditions and soil properties (mainly soil pH) shape different soil microbial communities [59], thus influencing the potential of soil C sequestration; the niche specialization and differentiation of soil microorganisms have been discussed in previous studies [60,61]. Except for the role of soil microorganisms, different sites may exhibit different interactions between SOM and the soil minerals, also involving the role of positively charged cations, such as Ca^{2+} , which acts as a bridge between negatively charged soil particles and/or organic molecules inducing SOM stabilization [62].

4.4. Identifying Soil and Plant Parameters Representing Best Soil Organic C Sequestration

In our study, POXC was used as an indicator of C sequestration following the general pattern of SOM; we found no significant yet positive relationship between POXC and SOM (Table 2). However, the latter agrees with previous findings showing a relation between POXC and SOM stabilization [63,64], suggesting that POXC could be a valid candidate indicator of soil C sequestration in semi-arid agroecosystems, such as those of the Mediterranean region. Furthermore, the SOM showed a positive relationship with TKN, as shown previously [7,65,66], revealing a solid coupling between C and N cycles in agroecosystems. By contrast, the CO_2 rate showed was not correlated to SOM, POCX, TKN, or another parameter (Table 2 and Figure 11) and did not vary significantly between tree cropping systems and sites, suggesting that a combination of factors may have been involved, such as the quality of SOM and the potential interactions with soil microorganisms, solid phase, and ions [62]. This interaction is perhaps the reason behind the relatively weak coupling between SOM and soil texture found in our study, which is opposite to previous studies [67,68]. Finally, in our study, the leaf C:N ratio had a positive relationship with the SOM, suggesting these tree parameters are relevant indicators of soil organic C sequestration, discussed above.

5. Conclusions

Based on the findings of the study, we conclude:

1. Tree cropping systems (olive groves and orange orchards) can enhance organic C stock in the soil under the semi-arid soil-climatic conditions of Crete, Greece.
2. Tree crops have different soil organic C sequestration potentials related to crop litter characteristics (e.g., leaf C:N ratio) and rhizosphere regulation. Olive trees showed higher soil organic C sequestration potential than orange trees.
3. SOM is challenging to be described by typical soil properties due to the synergetic effect of a plethora of abiotic and biotic parameters; in our study, SOM had a positive relationship with TKN (and less with POXC) in the soil as well as with the C:N ratio of the tree litter.
4. Further work should involve a more extensive survey and a more comprehensive range of plant and soil physicochemical (e.g., soil fractions) and microbial (e.g., microbial composition and structure) and biochemical soil parameters to identify the most critical ones to soil C sequestration. Another area of future research is the investigation of the coupling between the C and N cycles in semi-arid agroecosystems.

Author Contributions: Conceptualization, V.A.T.; methodology, V.A.T., E.K., N.M. and T.C.; software, N.M.; formal analysis, N.M.; investigation, E.S.; resources, E.K., V.A.T. and N.M.; data curation, N.M.; writing—original draft preparation, E.S. and V.A.T.; writing—review and editing, N.M., I.K., G.G., T.C. and E.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research was partially funded by the European Union LIFE+ Nature and Biodiversity project Improvement of green infrastructure (GI) in agroecosystems: reconnecting natural areas by countering habitat fragmentation—LIFE IGIC (Grant number LIFE16 NAT/GR/000575).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available upon request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Lal, R.; Negassa, W.; Lorenz, K. Carbon sequestration in soil. *Curr. Opin. Environ. Sustain.* **2015**, *15*, 79–86. [\[CrossRef\]](#)
2. Cramer, W.; Guiot, J.; Fader, M.; Garrabou, J.; Gattuso, J.-P.; Iglesias, A.; Lange, M.A.; Lionello, P.; Llasat, M.C.; Paz, S. Climate change and interconnected risks to sustainable development in the Mediterranean. *Nat. Clim. Change* **2018**, *8*, 972–980. [\[CrossRef\]](#)
3. Aguilera, E.; Diaz-Gaona, C.; Garcia-Laureano, R.; Reyes-Palomo, C.; Guzmán, G.I.; Ortolani, L.; Sanchez-Rodriguez, M.; Rodriguez-Estevez, V. Agroecology for adaptation to climate change and resource depletion in the Mediterranean region. A review. *Agric. Syst.* **2020**, *181*, 102809. [\[CrossRef\]](#)
4. Fisher, J.B.; Melton, F.; Middleton, E.; Hain, C.; Anderson, M.; Allen, R.; McCabe, M.F.; Hook, S.; Baldocchi, D.; Townsend, P.A. The future of evapotranspiration: Global requirements for ecosystem functioning, carbon and climate feedbacks, agricultural management, and water resources. *Water Resour. Res.* **2017**, *53*, 2618–2626. [\[CrossRef\]](#)
5. Banerjee, A.; Meena, R.S.; Jhariya, M.K.; Yadav, D.K. *Agroecological Footprints Management for Sustainable Food System*; Springer: Berlin, Germany, 2021.
6. Meena, R.S.; Kumar, S.; Yadav, G.S. Soil carbon sequestration in crop production. In *Nutrient Dynamics for Sustainable Crop Production*; Springer: Berlin, Germany, 2020; pp. 1–39.
7. Paranychanakis, N.V.; Giannakis, G.; Moraetis, D.; Tzanakakis, V.A.; Nikolaidis, N.P. Crop litter has a strong effect on soil organic matter sequestration in semi-arid environments. *Sustainability* **2021**, *13*, 13278. [\[CrossRef\]](#)
8. Pausch, J.; Kuzyakov, Y. Carbon input by roots into the soil: Quantification of rhizodeposition from root to ecosystem scale. *Glob. Change Biol.* **2018**, *24*, 1–12. [\[CrossRef\]](#)
9. De Deyn, G.B.; Cornelissen, J.H.; Bardgett, R.D. Plant functional traits and soil carbon sequestration in contrasting biomes. *Ecol. Lett.* **2008**, *11*, 516–531. [\[CrossRef\]](#)
10. Kallenbach, C.M.; Rolston, D.E.; Horwath, W.R. Cover cropping affects soil N₂O and CO₂ emissions differently depending on type of irrigation. *Agric. Ecosyst. Environ.* **2010**, *137*, 251–260. [\[CrossRef\]](#)
11. Badagliacca, G.; Ruisi, P.; Rees, R.M.; Saia, S. An assessment of factors controlling N₂O and CO₂ emissions from crop residues using different measurement approaches. *Biol. Fertil. Soils* **2017**, *53*, 547–561. [\[CrossRef\]](#)
12. Wilson, H.; Al-Kaisi, M. Crop rotation and nitrogen fertilization effect on soil CO₂ emissions in central Iowa. *Appl. Soil Ecol.* **2008**, *39*, 264–270. [\[CrossRef\]](#)
13. Sun, W.; Canadell, J.G.; Yu, L.; Yu, L.; Zhang, W.; Smith, P.; Fischer, T.; Huang, Y. Climate drives global soil carbon sequestration and crop yield changes under conservation agriculture. *Glob. Change Biol.* **2020**, *26*, 3325–3335. [\[CrossRef\]](#)
14. Jarecki, M.K.; Lal, R. Crop management for soil carbon sequestration. *Crit. Rev. Plant Sci.* **2003**, *22*, 471–502. [\[CrossRef\]](#)
15. Langgut, D.; Cheddadi, R.; Carrión, J.S.; Cavanagh, M.; Colombaroli, D.; Eastwood, W.J.; Greenberg, R.; Litt, T.; Mercuri, A.M.; Miebach, A. The origin and spread of olive cultivation in the Mediterranean Basin: The fossil pollen evidence. *Holocene* **2019**, *29*, 902–922. [\[CrossRef\]](#)
16. Proietti, S.; Sdringola, P.; Desideri, U.; Zepparelli, F.; Brunori, A.; Ilarioni, L.; Nasini, L.; Regni, L.; Proietti, P. Carbon footprint of an olive tree grove. *Appl. Energy* **2014**, *127*, 115–124. [\[CrossRef\]](#)
17. Sofo, A.; Nuzzo, V.; Palese, A.M.; Xiloyannis, C.; Celano, G.; Zukowskyj, P.; Dichio, B. Net CO₂ storage in Mediterranean olive and peach orchards. *Sci. Hortic.* **2005**, *107*, 17–24. [\[CrossRef\]](#)
18. Testi, L.; Orgaz, F.; Villalobos, F. Carbon exchange and water use efficiency of a growing, irrigated olive orchard. *Environ. Exp. Bot.* **2008**, *63*, 168–177. [\[CrossRef\]](#)
19. Villalobos, F.; Testi, L.; Hidalgo, J.; Pastor, M.; Orgaz, F. Modelling potential growth and yield of olive (*Olea europaea* L.) canopies. *Eur. J. Agron.* **2006**, *24*, 296–303. [\[CrossRef\]](#)
20. López-Bellido, R.J.; Fontán, J.M.; López-Bellido, F.J.; López-Bellido, L. Carbon sequestration by tillage, rotation, and nitrogen fertilization in a Mediterranean Vertisol. *Agron. J.* **2010**, *102*, 310–318. [\[CrossRef\]](#)

21. Zapata-Sierra, A.J.; Manzano-Agugliaro, F. Controlled deficit irrigation for orange trees in Mediterranean countries. *J. Clean. Prod.* **2017**, *162*, 130–140. [\[CrossRef\]](#)
22. Yang, S.; Aydin, M.; Kitamura, Y.; Yano, T. The impact of irrigation water quality on water uptake by orange trees. *Afr. J. Agric. Res.* **2010**, *5*, 2661–2667.
23. Germana, C.; Sardo, V. Determining when to initiate irrigation of orange trees. In Proceedings of the IV International Symposium on Irrigation of Horticultural Crops 664, Davis, CA, USA, 1–6 September 2003; pp. 591–597.
24. Consoli, S.; Stagno, F.; Roccuzzo, G.; Cirelli, G.; Intrigliolo, F. Sustainable management of limited water resources in a young orange orchard. *Agric. Water Manag.* **2014**, *132*, 60–68. [\[CrossRef\]](#)
25. Consoli, S.; Stagno, F.; Vanella, D.; Boaga, J.; Cassiani, G.; Roccuzzo, G. Partial root-zone drying irrigation in orange orchards: Effects on water use and crop production characteristics. *Eur. J. Agron.* **2017**, *82*, 190–202. [\[CrossRef\]](#)
26. Tsanis, I.K.; Koutroulis, A.G.; Daliakopoulos, I.N.; Jacob, D. Severe climate-induced water shortage and extremes in Crete. *Clim. Chang.* **2011**, *106*, 667–677. [\[CrossRef\]](#)
27. Tzanakakis, V.; Angelakis, A.; Paranychiakis, N.; Dialynas, Y.; Tchobanoglous, G. Challenges and Opportunities for Sustainable Management of Water Resources in the Island of Crete, Greece. *Water* **2020**, *12*, 1538. [\[CrossRef\]](#)
28. Allen, H.; Randall, R.; Amable, G.; Devereux, B. The impact of changing olive cultivation practices on the ground flora of olive groves in the Messara and Psiloritis regions, Crete, Greece. *Land Degrad. Dev.* **2006**, *17*, 249–273. [\[CrossRef\]](#)
29. Goward, J.; Whitty, M.; Ch, R. Estimating and predicting carbon sequestered in a vineyard with soil surveys, spatial data and G.I.S. management. Bachelor's Thesis, Engineering University of New South Wales, Kensington, Australia, 2012.
30. Lal, R. Soil carbon stocks under present and future climate with specific reference to European ecoregions. *Nutr. Cycl. Agroecosystems* **2008**, *81*, 113–127. [\[CrossRef\]](#)
31. Proietti, P.; Sdringola, P.; Brunori, A.; Ilarioni, L.; Nasini, L.; Regni, L.; Pelleri, F.; Desideri, U.; Proietti, S. Assessment of carbon balance in intensive and extensive tree cultivation systems for oak, olive, poplar and walnut plantation. *J. Clean. Prod.* **2016**, *112*, 2613–2624. [\[CrossRef\]](#)
32. Nair, P.R.; Nair, V.D.; Kumar, B.M.; Showalter, J.M. Carbon sequestration in agroforestry systems. *Adv. Agron.* **2010**, *108*, 237–307.
33. Kourgialas, N.N.; Anyfanti, I.; Karatzas, G.P.; Dokou, Z. An integrated method for assessing drought prone areas—Water efficiency practices for a climate resilient Mediterranean agriculture. *Sci. Total Environ.* **2018**, *625*, 1290–1300. [\[CrossRef\]](#)
34. Tzoraki, O.; Kritsotakis, M.; Baltas, E. Spatial Water Use efficiency Index towards resource sustainability: Application in the island of Crete, Greece. *Int. J. Water Resour. Dev.* **2015**, *31*, 669–681. [\[CrossRef\]](#)
35. Bouyoucos, G.J. Hydrometer method improved for making particle size analyses of soils 1. *Agron. J.* **1962**, *54*, 464–465. [\[CrossRef\]](#)
36. Nelson, D.A.; Sommers, L.E. Total carbon, organic carbon, and organic matter. *Methods Soil Anal. Part 2 Chem. Microbiol. Prop.* **1983**, *9*, 539–579.
37. Bremner, J.M. Nitrogen-total. *Methods Soil Anal. Part 3 Chem. Methods* **1996**, *5*, 1085–1121.
38. Weil, R.R.; Islam, K.R.; Stine, M.A.; Gruver, J.B.; Samson-Liebig, S.E. Estimating active carbon for soil quality assessment: A simplified method for laboratory and field use. *Am. J. Altern. Agric.* **2003**, *18*, 3–17.
39. Allison, S.D.; Jastrow, J.D. Activities of extracellular enzymes in physically isolated fractions of restored grassland soils. *Soil Biol. Biochem.* **2006**, *38*, 3245–3256. [\[CrossRef\]](#)
40. Bottomley, P.J.; Angle, J.S.; Weaver, R. *Methods of Soil Analysis, Part 2: Microbiological and Biochemical Properties*; John Wiley & Sons: New York, NY, USA, 2020; Volume 12.
41. Kapagianni, P.D.; Topalis, I.; Gwynn-Jones, D.; Menkissoglu-Spiroudi, U.; Stamou, G.P.; Papatheodorou, E.M. Effects of plant invaders on rhizosphere microbial attributes depend on plant identity and growth stage. *Soil Res.* **2020**, *59*, 225–238. [\[CrossRef\]](#)
42. Garcia-Franco, N.; Wiesmeier, M.; Collocho Hurtarte, L.C.; Fella, F.; Martínez-Mena, M.; Almagro, M.; Martínez, E.G.; Kögel-Knabner, I. Pruning residues incorporation and reduced tillage improve soil organic matter stabilization and structure of salt-affected soils in a semi-arid Citrus tree orchard. *Soil Tillage Res.* **2021**, *213*, 105129. [\[CrossRef\]](#)
43. Paterson, E.; Gebbing, T.; Abel, C.; Sim, A.; Telfer, G. Rhizodeposition shapes rhizosphere microbial community structure in organic soil. *New Phytol.* **2007**, *173*, 600–610. [\[CrossRef\]](#)
44. Benizri, E.; Dedourge, O.; Dibattista-Leboeuf, C.; Piutti, S.; Nguyen, C.; Guckert, A. Effect of maize rhizodeposits on soil microbial community structure. *Appl. Soil Ecol.* **2002**, *21*, 261–265. [\[CrossRef\]](#)
45. Semchenko, M.; Xue, P.; Leigh, T. Functional diversity and identity of plant genotypes regulate rhizodeposition and soil microbial activity. *New Phytol.* **2021**, *232*, 776–787. [\[CrossRef\]](#)
46. Tamura, M.; Suseela, V.; Simpson, M.; Powell, B.; Tharayil, N. Plant litter chemistry alters the content and composition of organic carbon associated with soil mineral and aggregate fractions in invaded ecosystems. *Glob. Chang. Biol.* **2017**, *23*, 4002–4018. [\[CrossRef\]](#)
47. Castellano, M.J.; Mueller, K.E.; Olk, D.C.; Sawyer, J.E.; Six, J. Integrating plant litter quality, soil organic matter stabilization, and the carbon saturation concept. *Glob. Chang. Biol.* **2015**, *21*, 3200–3209. [\[CrossRef\]](#)
48. Redin, M.; Guénon, R.; Recous, S.; Schmatz, R.; de Freitas, L.L.; Aita, C.; Giacomini, S.J. Carbon mineralization in soil of roots from twenty crop species, as affected by their chemical composition and botanical family. *Plant Soil* **2014**, *378*, 205–214. [\[CrossRef\]](#)
49. Talbot, J.M.; Yelle, D.J.; Nowick, J.; Treseder, K.K. Litter decay rates are determined by lignin chemistry. *Biogeochemistry* **2012**, *108*, 279–295. [\[CrossRef\]](#)

50. Forney, D.C.; Rothman, D.H. Common structure in the heterogeneity of plant-matter decay. *J. R. Soc. Interface* **2012**, *9*, 2255–2267. [\[CrossRef\]](#)
51. Rahman, M.M.; Tsukamoto, J.; Rahman, M.M.; Yoneyama, A.; Mostafa, K.M. Lignin and its effects on litter decomposition in forest ecosystems. *Chem. Ecol.* **2013**, *29*, 540–553. [\[CrossRef\]](#)
52. Giannakis, G.V.; Panakoulia, S.K.; Nikolaidis, N.P.; Paranychanakis, N.V. Simulating Soil Fertility Restoration Using the CAST Model. *Procedia Earth Planet. Sci.* **2014**, *10*, 325–329. [\[CrossRef\]](#)
53. Nieto, O.M.; Castro, J.; Fernández, E.; Smith, P. Simulation of soil organic carbon stocks in a Mediterranean olive grove under different soil-management systems using the RothC model. *Soil Use Manag.* **2010**, *26*, 118–125. [\[CrossRef\]](#)
54. Tzanakakis, V.A.; Paranychanakis, N.V. Divergent response of ammonia oxidizers to various amino acids. *Appl. Soil Ecol.* **2017**, *114*, 45–51. [\[CrossRef\]](#)
55. Qiu, H.; Ge, T.; Liu, J.; Chen, X.; Hu, Y.; Wu, J.; Su, Y.; Kuzyakov, Y. Effects of biotic and abiotic factors on soil organic matter mineralization: Experiments and structural modeling analysis. *Eur. J. Soil Biol.* **2018**, *84*, 27–34. [\[CrossRef\]](#)
56. Cai, A.; Feng, W.; Zhang, W.; Xu, M. Climate, soil texture, and soil types affect the contributions of fine-fraction-stabilized carbon to total soil organic carbon in different land uses across China. *J. Environ. Manag.* **2016**, *172*, 2–9. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Angst, G.; Messinger, J.; Greiner, M.; Häusler, W.; Hertel, D.; Kirfel, K.; Kögel-Knabner, I.; Leuschner, C.; Rethemeyer, J.; Mueller, C.W. Soil organic carbon stocks in topsoil and subsoil controlled by parent material, carbon input in the rhizosphere, and microbial-derived compounds. *Soil Biol. Biochem.* **2018**, *122*, 19–30. [\[CrossRef\]](#)
58. Moinet, G.Y.; Hunt, J.E.; Kirschbaum, M.U.; Morcom, C.P.; Midwood, A.J.; Millard, P. The temperature sensitivity of soil organic matter decomposition is constrained by microbial access to substrates. *Soil Biol. Biochem.* **2018**, *116*, 333–339. [\[CrossRef\]](#)
59. Goss-Souza, D.; Mendes, L.W.; Borges, C.D.; Baretta, D.; Tsai, S.M.; Rodrigues, J.L. Soil microbial community dynamics and assembly under long-term land use change. *FEMS Microbiol. Ecol.* **2017**, *93*, fix109. [\[CrossRef\]](#) [\[PubMed\]](#)
60. Lennon, J.T.; Aanderud, Z.T.; Lehmkuhl, B.K.; Schoolmaster, D.R., Jr. Mapping the niche space of soil microorganisms using taxonomy and traits. *Ecology* **2012**, *93*, 1867–1879. [\[CrossRef\]](#)
61. Prosser, J.I.; Nicol, G.W. Archaeal and bacterial ammonia-oxidisers in soil: The quest for niche specialisation and differentiation. *Trends Microbiol.* **2012**, *20*, 523–531. [\[CrossRef\]](#)
62. Mikutta, R.; Kleber, M.; Torn, M.S.; Jahn, R. Stabilization of soil organic matter: Association with minerals or chemical recalcitrance? *Biogeochemistry* **2006**, *77*, 25–56. [\[CrossRef\]](#)
63. Hurisso, T.T.; Culman, S.W.; Horwath, W.R.; Wade, J.; Cass, D.; Beniston, J.W.; Bowles, T.M.; Grandy, A.S.; Franzluebbers, A.J.; Schipanski, M.E.; et al. Comparison of Permanganate-Oxidizable Carbon and Mineralizable Carbon for Assessment of Organic Matter Stabilization and Mineralization. *Soil Sci. Soc. Am. J.* **2016**, *80*, 1352–1364. [\[CrossRef\]](#)
64. Sprunger, C.D.; Martin, T.; Mann, M. Systems with greater perenniality and crop diversity enhance soil biological health. *Agric. Environ. Lett.* **2020**, *5*, e20030. [\[CrossRef\]](#)
65. Yao, Z.; Zhang, D.; Liu, N.; Yao, P.; Zhao, N.; Li, Y.; Zhang, S.; Zhai, B.; Huang, D.; Wang, Z.; et al. Dynamics and Sequestration Potential of Soil Organic Carbon and Total Nitrogen Stocks of Leguminous Green Manure-Based Cropping Systems on the Loess Plateau of China. *Soil Tillage Res.* **2019**, *191*, 108–116. [\[CrossRef\]](#)
66. Tong, C.; Xiao, H.; Tang, G.; Wang, H.; Huang, T.; Xia, H.; Keith, S.J.; Li, Y.; Liu, S.; Wu, J. Long-term fertilizer effects on organic carbon and total nitrogen and coupling relationships of C and N in paddy soils in subtropical China. *Soil Tillage Res.* **2009**, *106*, 8–14. [\[CrossRef\]](#)
67. Hamarashid, N.H.; Othman, M.A.; Hussain, M.-A.H. Effects of soil texture on chemical compositions, microbial populations and carbon mineralization in soil. *Egypt. J. Exp. Biol.(Bot.)* **2010**, *6*, 59–64.
68. Augustin, C.; Cihacek, L.J. Relationships between soil carbon and soil texture in the Northern Great Plains. *Soil Sci.* **2016**, *181*, 386–392. [\[CrossRef\]](#)

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.